



Review article

Molecular chaperones and the aging process

Csaba Sőti & Péter Csermely*

Department of Medical Chemistry, Semmelweis University, P.O. Box 260, Budapest 8, H-1444, Hungary;

*Author for correspondence (e-mail: csermely@puskin.sote.hu; fax: +36-1-266-6550)

Received 14 February 2000; accepted 23 February 2000

Key words: aging, Alzheimer's disease, chaperone, heat shock protein, immune response, longevity, neurodegenerative diseases, protein folding, senescence, stress protein

Abstract

Molecular chaperones are abundant, well-conserved proteins responsible for the maintenance of the conformational homeostasis of cellular proteins and RNAs. Environmental stress is a proteotoxic insult to the cell, which leads to chaperone (heat shock protein, stress protein) induction. The protective role of chaperones is a key factor for cell survival and in repairing cellular damage. The present review summarizes our current knowledge about changes in chaperone expression and function in the aging process, as well as their possible involvement in the development of longevity and cellular senescence. We also overview their putative role in neurodegenerative diseases, such as in Alzheimer's disease and the changes in immune and autoimmune response against various chaperones in aging.

Abbreviations: Grp – glucose regulated protein; Hsc70 – the non-inducible (cognate) form of the 70 kDa heat shock protein; Hsp – heat shock protein

Introduction: cellular roles of molecular chaperones

Molecular chaperones bind to, and stabilize an otherwise unstable conformer of another protein or RNA and, by controlled binding and release, facilitate its correct fate *in vivo*: be it folding, oligomeric assembly, transport to a particular subcellular compartment, or disposal by degradation. In the molecular level chaperones (1) protect against aggregation, (2) solubilize protein aggregates, (3) assist in protein folding/refolding by partial unfolding the intermediate structures in the folding process, (3) target ultimately damaged proteins to degradation and (5) sequester overloaded damaged proteins to larger aggregates. Chaperones are ubiquitous, highly conserved proteins, which probably played a major role in the evolution of modern enzymes (Csermely 1997, 1999; Hartl 1996). Chaperones are vital for our cells during their whole lifetime. However, they are needed even more

after environmental stress, that induces protein damage. Stress (especially its most studied archetype, heat shock) leads to the expression of most chaperones, which therefore are called heat-shock, or stress proteins. Lacking a settled view about their action in the molecular level, chaperones are still best classified by their molecular weights. The major chaperone families are listed in Table 1. Besides the general chaperones listed in Table 1, which have a rather large promiscuity in target-selection, there are also specialized chaperones, such as Hsp47, the special chaperone of collagen. Chaperones usually do not increase the speed of protein folding, just inversely, by binding to folding intermediates, or by their repetitive pulling attempts extend the total folding time and simultaneously increase the final yield of the native protein. Special steps of protein folding are accelerated by the so-called 'folding catalysts', such as peptidyl-prolyl isomerases (immunophilins), and protein disulfide isomerases, which promote the cis-trans isomerization

Table 1. Major molecular chaperone families.

Eukaryotic chaperone family members ^a	Recent reviews
Hsp25, Hsp27, crystallins, small heat-shock proteins	Arrigo 1998; Derham and Harding 1999
Hsp60, chaperonins	Bukau and Horwich 1998; Hartl 1996
Hsp70, Grp78	Bukau and Horwich 1998; Hartl 1996
Hsp90, Grp94	Buchner 1999; Csermely et al. 1998; Pratt and Toft 1997
Hsp104	Schirmer et al. 1996

^aNeither the co-chaperones (chaperones which help the function of other chaperones listed, such as Hsp10, Hsp40, Hip, Hop, Hup, etc.), nor the so-called folding catalysts, the peptidyl-prolyl isomerases (immunophilins) and protein disulfide isomerases were included in this table, albeit almost all of these proteins also possess a 'traditional' chaperone activity in their own right.

of peptide bonds adjacent to proline residues and the formation of disulfide bridges in the endoplasmic reticulum, respectively.

Besides the need for assisted folding of *de novo* synthesized, nascent proteins, and the chaperone-mediated repair of protein damage, cellular life requires a constant remodelling of cell structure. Chaperones provide an essential help in all these processes. Individual chaperone proteins usually do not work alone, but either form a highly structured homo- or heterooligomeric complex, such as the Hsp60 chaperone machine, or assemble to a dynamic cohort of chaperones such as the Hsp90-containing foldosome in the cytoplasm or its counterpart in the endoplasmic reticulum. Most major chaperones are helped by several smaller co-chaperones (such as Hsp10 in case of Hsp60, Hsp40 in case of Hsp70 and p23 in case of Hsp90). The large and dynamic assemblies of individual chaperones are attached to the microfilament and microtubular system. About twenty years ago based on high-voltage electron microscopy Porter and co-workers suggested the existence of a cellular meshwork, called as 'microtrabecular lattice' to organize cytoplasmic proteins and RNA-s (Wolosewick and Porter 1979; Schliwa et al. 1981). Some later reports questioned their original experiments and considered the lattice as an artifact of the techniques used. However, more and more data provide indirect evidence for a high-order organization of the cytoplasm. Chaperones are ideal candidates for being a major constituent of a cytoplasmic meshwork (Csermely et al. 1998; Pratt and Toft 1997).

Protein folding and the aging process

During the life-span of a stable protein various posttranslational modifications occur (Harding et al.

1989). These include deamidation of asparaginyl and glutaminyl residues and the subsequent formation of isopeptide bonds (Wright 1991), protein glycation, methionine oxidation (Sun et al. 1999), etc. Susceptibility to oxidative damage differs protein to protein, as has been demonstrated in a comparison of bovine serum albumin and glutamine synthase (Berlett et al. 1996), and of various K⁺ channels (Duprat et al., 1995), suggesting a role for different tertiary-quaternary structure as well as folding. Even in early studies aging-induced inactivation of isocitrate-lyase (Reiss and Rothstein 1974), or phosphoglycerate kinase (Yuh and Gafni 1987) could be associated with the accumulation of a non-native, heat labile conformation of the enzymes. In a refolding study the increased helical content of 'old' aldolase was preserved after refolding of the enzyme, which suggested that the conformational changes were mostly induced by the various posttranslational modifications during the life of the protein (Demchenko et al. 1983).

The ocular lens is a transparent organ comprised of a highly concentrated and ordered matrix of structural proteins, called crystallins, which are probably the longest lived proteins of the body. Lens transparency is dependent upon maintenance of the short range order of the crystalline matrix. The gradual loss of this order leads to the opacification of the lens and in extreme cases to the development of cataract. Therefore crystallins became excellent substrates of widespread research activity in determining various posttranslational modification of aged proteins. Besides glycation, methionine oxidation and crystallin-crystallin crosslinking (Sharma et al. 1995; Smith et al. 1997), a loss of N-terminal amino acids, covalent modification of C-terminal lysine and photodegradation of crystallin tryptophanes were also reported (Lin et al. 1997; Kamei et al. 1997; Schauerte and Gafni 1995). Aging α -crystallin also undergoes

Table 2. Changes of chaperone expression in aging.

Chaperone	Change	References
<i>Chaperone levels</i>		
Hsp22, Hsp23, Hsp70	Elevated in <i>Drosophila</i>	Wheeler et al. 1995
Hsp47, Hsp70	Elevated in rat kidneys	Maiello et al. 1998; Razzaque et al. 1998
Hsc70 ^a	No change in hepatocytes	Wu et al. 1993
Hsc70 ^a	Decrease in testis	Krawczyk et al. 1989
<i>Chaperone induction</i>		
Hsp27	Heat induction is impaired	Rao et al. 1999
Heme oxygenase (Hsp32)	Oxygen damage induction is impaired	Nakanishi and Yasumoto, 1997
Superoxide dismutase	Heat induction is impaired	Niedzwiecki et al. 1992
Hsp60	Heat induction is impaired	Rao et al. 1999
Hsp70	Heat, ischemia and mitogen induction are impaired	Blake et al. 1991; Deguchi et al. 1988; Faassen et al. 1988; Fargnoli et al. 1990; Heydari et al. 1994; Liu et al. 1996; Niedzwiecki et al. 1991; Nitta et al. 1994 ^b
Hsp70	Exercise induction is maintained	Kregel and Moseley 1996

^aHsc70 denotes the non-inducible (cognate) form of Hsp70.

^bThe large number of reports allowed us to cite only those, which were among the firsts, or provided a review of other studies.

a self-cleavage at Asn-101 following the formation of the succinimide intermediate of the deamidation reaction (Voorter et al. 1988).

Thus a large variety of posttranslational modifications accumulate in proteins having an extended life-span. Although in normal conditions intracellular protein turnover is rather fast, carbonyl content of aging proteins – an indicator of oxidative damage – increases threefold in several tissues, such as brain, heart and liver (Stadtman 1992). However, extracellular proteins, like collagen also have a longer lifetime, and their accumulating proteotoxic damage leads to increased tissue rigidity as well as impairs cell–cell communication (Monnier and Cerami 1981).

Protein degradation is mostly accomplished by the proteasome and helped by various chaperones. Aging leads to a decrease in adaptive responses and consequently to an increased occurrence of proteotoxic conditions inside the cell. Aging also attenuates both the ‘cellular surveillance’ of chaperones to recognize damaged proteins, and the activity of the major cytoplasmic proteolytic apparatus, the proteasome (Conconi et al. 1996; Heydari et al. 1994; Liu et al. 1996). Besides the decline in the activation of protease systems, some oxidized, crosslinked proteins are much poorer substrates, but highly effective inhibitors of the proteasome (Friguet et al. 1994). All these events cause a massive accumulation of posttranslationally modified, misfolded proteins.

Chaperones and aging

The accumulation of misfolded proteins in aged organisms would require an increased amount of chaperones to prevent protein aggregation and to assist in refolding, or degradation. This may be the reason, why some aged species develop a constitutively increased level of several chaperones, such as Hsp22, or Hsp70 (Table 2). The higher amount of chaperone proteins is especially characteristic to the adaptive response of aging kidney, where increased fibrosis requires an additional amount of the collagen-specific Hsp47 (Razzaque et al. 1998). On the other hand, there is a large number of reports demonstrating that the induction of various chaperones is impaired in aged organisms (Table 2). Interestingly, while heat-induced synthesis of Hsp70 is impaired in aged rats, exercise in the same animal is able to induce a significant amount of Hsp70 (Kregel and Moseley 1996). Different changes in the induction mechanisms of various chaperones during the aging process are further substantiated by the findings of Fleming et al. (1988), who showed a substantially altered pattern of heat shock protein induction in old fruit flies compared to young species.

Differences in chaperone induction in aged animals and human subjects (Table 2) exclude the possibility of a general impairment in the transcriptional process of molecular chaperones. Indeed, the

level of heat shock factor-1, the transcription factor responsible for the induction of most chaperones is practically unchanged during aging. However, activation and binding of heat shock factor-1 to the heat shock element, its DNA-binding site in the promoter region of molecular chaperones is decreased in aged animals (Heydari et al. 1994; Pahlavani et al. 1995; Locke and Tanguay 1996). The exact mechanism of the defective activation is not known. In recent years several heat shock factor-binding proteins were identified, which all modulate the heat shock response (Morimoto 1998), and may well constitute the molecular mechanism of the differential impairment of chaperone-induction during aging.

Interestingly, there are much less reports on the functional changes of molecular chaperones during aging than those on their level, or induction. Chaperone activity of alpha crystallin is markedly decreased in senile human lenses (Cherian et al. 1995; Derham and Harding 1997). Due to the lack of protein synthesis and degradation in the lens, crystallins are one of the longest lived proteins in the human body, and therefore they are especially prone to various proteotoxic damage (see above). Hence it is not surprising that their activity is diminished in senescent lenses. This situation makes the so far not addressed question even more exciting, whether an impaired activity of other chaperones might also add to the damage caused by their diminished induction in aged organisms. As another of the sporadic examples of chaperone function in aged animals or human subjects, Hsp90 protects the age-related decline of proteasome activity. However, the association of Hsp90 with the proteasome decreases with age which may lead to an enhanced vulnerability of the proteasome for stress-induced damage in aged organisms (Conconi et al. 1996).

Chaperones in neurodegenerative diseases

Accumulation of misfolded proteins in aged organisms is especially pronounced in postmitotic cells, such as in neurons. The threat of damaged proteins becomes even greater if the protein is protease-resistant. The difficulties of protein degradation together with an impaired protease activity and chaperone action in aging neurons, lead to a massive accumulation of these proteins, and causes neurodegeneration. The best known example of this propagating disease is Alzheimer's disease, however several

other neurodegenerative diseases, such as prion disease, Huntington disease or others may also be mentioned. Several studies showed the induction of small heat shock proteins (Hsp27, crystallin), Hsp70 and ubiquitin (a 6 kDa heat shock protein, which labels damaged proteins and directs them for proteolytic degradation) in neurons affected by Alzheimer's disease and in surrounding astrocytes. Neuronal chaperones were localized in neuritic plaques and neurofibrillary tangles and were probably participating in the heroic attempts of the affected neuron to sequester the beta-amyloid and other damaged proteins (Cisse et al. 1993; Hamos et al. 1991; Perez et al. 1991; Renkawek et al. 1993; Shinohara et al. 1993). Interestingly the endoplasmic homologue of Hsp70, Grp78 showed an increased expression within successfully surviving neurons (Hamos et al. 1991). Other, not affected cells of Alzheimer victims, such as olfactory neurons (Getchell et al. 1995) or mononuclear blood cells (Wakutani et al. 1995) showed a decreased expression of Hsp70.

Chaperones and cellular senescence

Fibroblasts and other freshly isolated cells undergo only a limited number of divisions in cell culture. During the consequent duplications these cells change many of their original properties, a process, which is called cellular senescence (Smith and Pereira-Smith 1996). Increasing cellular senescence – on one hand – is well correlated with organismal aging. On the other hand, its induction by oncogenic stimuli contributes to organismal longevity by reducing the occurrence of cancer (Serrano et al. 1997). Cellular aging of fibroblasts is known to impair the induction of several chaperones, such as the collagen-specific Hsp47 (Miyashi et al. 1995), Hsp70 and Hsp90 (Liu et al. 1989; Cristofalo et al. 1989). Similarly to the mechanism found in aged animals, activation and binding of heat shock factor-1 to the heat shock element is decreased in aged cells (Choi et al. 1990; Effros et al. 1994). The exact mechanism of the defective activation is also not known. In some studies a decrease in the amount of heat shock factor-1 has been found (Gutssmann-Conrad et al. 1998), while other studies suggest the presence of an inhibitory compound (Choi et al. 1990). In a recent report Bonelli et al. (1999) found an impairment in the posttranslational processing of Hsp70 mRNA resulting in its impaired nuclear export.

The defect of aged cells to induce Hsp70 may lead to their death in an unexpected way. Hsp70 is known to mediate the suppression of a stress-activated kinase, JNK, an early component of stress-induced apoptotic signalling pathway. Lacking a proper activation of Hsp70 senescent cells became prone for accelerated apoptosis after various stressful stimuli, such as heat shock (Gabai et al. 1998).

Several chaperones have a direct effect on cellular senescence. Overexpression of Hsp27 in bovine arterial endothelial cells leads to an accelerated growth and senescence. Interestingly, when a mutant, nonphosphorylatable form of Hsp27 was expressed, cellular senescence was hindered (Piotrowicz et al. 1995). When a mortality factor has been isolated from cytoplasmic extracts of senescing (mortal) fibroblasts, it turned to be a member of the Hsp70 chaperone family. This Hsp70-homologue, called mortalin is able to confer cellular senescence if transfected to immortal NIH 3T3 cells, and an antibody against mortalin could transiently stimulate cell division of senescent fibroblasts. Interestingly the protein exists in two isoforms, out of which only the cytosolic form is active, and its perinuclear homologue is not (Wadhwa et al. 1993a, b). As another possible involvement of chaperones in the regulation of cellular senescence, the 90 kDa heat shock protein, Hsp90 is required for the correct assembly and function of telomerase, a major enzyme involved in determining the life-span of cells (Holt et al. 1999).

Chaperones and longevity

Despite of the above accelerating effects of Hsp27 and the Hsp70 homologue, mortalin on senescence at the *cellular level in vitro*, there are several reports suggesting that increased chaperone action may also lead to an increased longevity of uni-, or multicellular *whole organisms in vivo*. Thermally conditioned *Drosophila* (Khazaeli et al. 1997) or *Caenorhabditis elegans* (Lithgow et al. 1995) exhibit greater longevity. Heat stress also induces an increased life-span of yeast (Shama et al. 1998), a process in which both Ras1 and Ras2, known to decrease and increase yeast life-span, respectively (Sun et al. 1994), and Hsp104, the key molecular chaperone inducing yeast thermotolerance were involved. Heat-shock induction of Hsp70 (Tatar et al. 1997), or overexpression of the heat shock protein, EF-1 alpha (Shepherd et al. 1989; Shikama et al. 1994) promotes fruit fly longev-

ity. Hsp70 induction was lower in the liver of aged mice prone to accelerated senescence than in mice that were resistant to accelerated senescence (Nakanishi et al. 1997). Moreover, a close correlation was found between stress resistance and longevity in several long-lived *Caenorhabditis elegans* and *Drosophila* mutants, which were either engineered genetically, or selected with classical population genetics (Lithgow and Kirkwood 1996). These examples confirm the hypothesis that a better adaptation capacity to various stresses make a major contribution to life span extension.

Caloric restriction is the only effective experimental manipulation known to retard aging in rodents, and this manipulation has been shown to alter a variety of processes that change with age including the oxidative damage of proteins (Sohal and Weindruch 1996; Youngman et al. 1992). Caloric restriction (60% of *ad libitum* diet causing a 43% increase in life span) increased the induction of Hsp70 of hepatocytes (Heydari et al. 1993), proximal gut (Ehrenfried et al. 1996), alveolar macrophages (Moore et al. 1998), but not of splenocytes (Pahlavani et al. 1996) of aged rats compared to their aged littermates being on *ad libitum* diet. In several cases caloric restriction parallel with an induction of an extended life span, restored the impaired chaperone-induction of aged animals to the 'young' level.

Changes in the immune response against chaperones in aging

Molecular chaperones are highly conserved proteins throughout the evolution. Bacteria and other infectious organisms experience a large stress when invade the human body. The general stress response is turned on, and they overexpress a wide variety of chaperone molecules. Several of the induced chaperones also appears on the surface of infectious bacteria and parasites. The immune system develops an immune response against these antigens during the first infection occurring in the very first days of postnatal development. This immune response becomes more and more robust, since the chaperone-antigens of various infectious organisms are highly similar to each other. In several cases, when a self-protein shares an epitop with a bacterial heat shock protein or the antibacterial immune response becomes misdirected, an autoimmune attack might develop.

How does this (auto)immune response change during the aging process? The answer is not simple. On one hand as the individual ages, the exposure to the infectious, or self-antigen becomes more manifest, the immune-memory gets stronger. This explains the findings, where anti-chaperone antibody levels were found to increase with increasing age, such as the anti-Hsp70 in human malaria infections (Alexandre et al. 1997), the anti-Hsp90 in human systemic lupus erythematosus (Faulds et al. 1995), or the rat anti-DnaK (the *Escherichia coli* homologue of Hsp70) immune response (Kimura et al. 1996). On the other hand, the general decline of immune responses in aged individuals may also impair the anti-chaperone immune response. This might be the explanation for the decline in anti-Hsp70 antibodies in dilated cardiomyopathy (Portig et al. 1997), or for the decrease in anti-synthetic peptide autoantigen antibodies (Marchalonis et al. 1993).

Perspectives and closing remarks

Aging can be defined as a multicausal process leading to a gradual decay of self defensive mechanisms, and an exponential accumulation of damage at the molecular-cellular and organismal level. The attenuation in molecular chaperone function and the simultaneous protein oxidation, misfolding and aggregation in aged organisms, as well as the correlation between adaptation capacity and life span raise the possibility that preservation of protein homeostasis and long-range protein organization can be major determinants in longevity (Figure 1). There are a plethora of opportunities to explore the underlying mechanisms behind these events.

- The exact mechanism of the attenuation of chaperone induction in aged organisms and in senescent cells will reveal interesting elements of the regulation of the stress response.
- There is a lot to do in examining the chaperone function in aged animals or cells, preferably using more complex, *in vivo* systems, such as expressed reporter proteins for folding, like luciferase. The role of chaperones in preventing membrane damage, or RNA-misfolding also deserves a much greater attention in aged organisms.
- The role of chaperones in cellular senescence, and in longevity is an area, which also keeps a lot of surprises for the coming years.

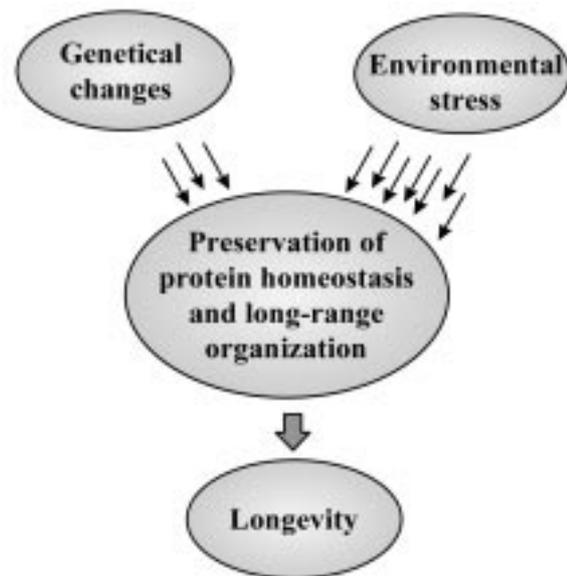


Figure 1. Hypothetical role of protein homeostasis and organization in longevity. Attenuation in molecular chaperone function and the simultaneous protein damage in aged organisms, as well as the correlation between stress adaptation capacity and life span raise the possibility that preservation of protein homeostasis and long-range protein organization can be major determinants in longevity. The putative role of chaperones in cytoplasmic protein organization is discussed in Pratt (1997) and in Csermely et al. (1998). The role of chaperones in buffering genetical changes providing a possibility for bursts in evolution has been recently demonstrated by Rutherford and Lindquist (1998).

Acknowledgements

Work in the authors' laboratory was supported by research grants from ICGEB, Hungarian Science Foundation (OTKA-T25206), Hungarian Ministry of Social Welfare (ETT-493/96), and the Volkswagen Foundation (I/73612). P.C. is an International Research Scholar of the Howard Hughes Medical Institute.

References

- Alexandre CO, Camargo LM, Mattei D et al. (1997) Humoral immune response to the 72 kDa heat shock protein from *Plasmodium falciparum* in populations at hypoendemic areas of malaria in western Brazilian Amazon. *Acta Trop* 64: 155–166
- Arrigo AP (1998) Small stress proteins: chaperones that act as regulators of intracellular redox state and programmed cell death. *Biol Chem* 379: 19–26
- Berlett BS, Levine RL and Stadtman ER (1996) Comparison of the effects of ozone on the modification of amino acid residues in glutamine synthase and bovine serum albumin. *J Biol Chem* 271: 4177–4182

- Blake MJ, Udelsman R, Feulner GJ et al. (1991) Stress-induced heat shock protein 70 expression in adrenal cortex: an adrenocorticotrophic hormone-sensitive, age-dependent response. *Proc Natl Acad Sci USA* 88: 9873–9877
- Bonelli MA, Alfieri RR, Petronini PG et al. (1999) Attenuated expression of 70-kDa heat shock protein in WI-38 human fibroblasts during aging *in vitro*. *Exp Cell Res* 252: 20–32
- Buchner J (1999) Hsp90 & co. – a holding for folding. *Trends Biochem Sci* 24: 136–141
- Bukau B and Horwich AL (1998) The Hsp70 and Hsp60 chaperone machines. *Cell* 92: 351–366
- Cherian M and Abraham EC (1995) Decreased molecular chaperone property of alpha-crystallins due to posttranslational modifications. *Biochem Biophys Res Commun* 208: 675–679
- Choi HS, Lin Z, Li BS et al. (1990) Age-dependent decrease in the heat-inducible DNA sequence-specific binding activity in human diploid fibroblasts. *J Biol Chem* 265: 18005–18011
- Cisse S, Perry G, Lacoste-Royal G et al. (1993) Immunochemical identification of ubiquitin and heat-shock proteins in corpora amylacea from normal aged and Alzheimer's disease brains. *Acta Neuropathol Berl* 85: 233–240
- Conconi M, Szewda LI, Levine RL et al. (1996) Age-related decline of rat liver multicatalytic proteinase activity and protection from oxidative inactivation by heat shock protein 90. *Arch Biochem Biophys* 331: 232–240
- Cristofalo VJ, Doggett DL, Brooks-Frederich KM et al. (1998) Growth factors as probes of cell aging. *Exp Gerontol* 24: 367–374
- Csermely P (1997) Proteins, RNA-s, chaperones and enzyme evolution: a folding perspective. *Trends Biochem Sci* 22: 147–149
- Csermely, P (1999) The 'chaperone-percolator' model: a possible molecular mechanism of Anfinsen-cage type chaperone action. *BioEssays* 21: 959–965
- Csermely P, Schnaider T, Söti Cs et al. (1998) The 90 kDa molecular chaperone family: structure, function and clinical applications. A comprehensive review. *Pharmacol Therapeutics* 79: 129–168
- Demchenko AP, Orlovskaya NN and Sukhomudrenko AG (1983) Age-dependent changes of protein structure. The properties of young and old rabbit aldolase are restored after reversible denaturation. *Exp Gerontol* 18: 437–446
- Deguchi Y, Negoro S and Kishimoto S (1988) Age-related changes of heat shock protein gene transcription in human peripheral blood mononuclear cells. *Biochem Biophys Res Commun* 157: 580–584
- Derham BK and Harding JJ (1997) Effect of aging on the chaperone-like function of human alpha-crystallin assessed by three methods. *Biochem J* 328: 763–768
- Derham BK and Harding JJ (1999) Alpha-crystallin as a molecular chaperone. *Progr Retinal Eye Res* 18: 463–509
- Duprat F, Guillemare E, Romey G et al. (1995) Susceptibility of cloned K⁺ channels to reactive oxygen species. *Proc Natl Acad Sci USA* 92: 11796–11800
- Effros RB, Zhu X and Walford RL (1994) Stress response of senescent T lymphocytes: reduced hsp70 is independent of the proliferative block. *J Gerontol* 49: B65–B70
- Ehrenfried JA, Evers BM, Chu KU et al. (1996) Caloric restriction increases the expression of heat shock protein in the gut. *Ann Surg* 223: 592–599
- Faassen AE, O'Leary JJ, Rodysill KJ et al. (1989) Diminished heat-shock protein synthesis following mitogen stimulation of lymphocytes from aged donors. *Exp Cell Res* 183: 326–334
- Fargnoli J, Kunisada T, Fornace AJ Jr et al. (1990) Decreased expression of heat shock protein 70 mRNA and protein after heat treatment in cells of aged rats. *Proc Natl Acad Sci USA* 87: 846–850
- Faulds G, Conroy S, Madaio M et al. (1995) Increased levels of antibodies to heat shock proteins with increasing age in Mrl/Mp-lpr/lpr mice. *Br J Rheumatol* 34: 610–615
- Fleming JE, Walton JK, Dubitsky R et al. (1988) Aging results in an unusual expression of *Drosophila* heat shock proteins. *Proc Natl Acad Sci USA* 85: 4099–4103
- Friguet B, Stadtman ER and Szewda LI (1994) Modification of glucose-6-phosphate dehydrogenase by 4-hydroxy-2-nonenal. Formation of cross-linked protein that inhibits the multicatalytic protease. *J Biol Chem* 269: 21639–21643
- Gabai VL, Meriin AB, Yaglom JA et al. (1998) Role of Hsp70 in regulation of stress-kinase JNK: implications in apoptosis and aging. *FEBS Lett* 438: 1–4
- Getchell TV, Krishna NS, Dhooper N et al. Human olfactory receptor neurons express heat shock protein 70: age-related trends. *Ann Otol Rhinol Laryngol* 104: 47–56
- Guttmann-Conrad A, Heydari AR, You S et al. (1998) The expression of heat shock protein 70 decreases with cellular senescence *in vitro* and in cells derived from young and old human subjects. *Exp Cell Res* 241: 404–413
- Hamos JE, Oblas B, Pulaski-Salo D et al. (1991) Expression of heat shock proteins in Alzheimer's disease. *Neurology* 41: 345–350
- Harding JJ, Beswick HT, Ajiboye R et al. (1989) Non-enzymatic post-translational modification of proteins in aging. A review. *Mech Aging Dev* 50: 7–16
- Hartl F-U (1996) Molecular chaperones in cellular protein folding. *Nature* 381: 571–580
- Heydari AR, Wu B, Takahashi R et al. (1993) Expression of heat shock protein 70 is altered by age and diet at the level of transcription. *Mol Cell Biol* 13: 2909–2918
- Heydari AR, Takahashi R, Gutsmann A et al. (1994) Hsp70 and aging. *Experientia* 50: 1092–1098
- Holt SE, Aisner DL, Baur J et al. (1999) Functional requirement of p23 and Hsp90 in telomerase complexes. *Genes Dev* 13: 817–826
- Kamei A, Iwase H and Masuda K (1997) Cleavage of amino acid residue(s) from the N-terminal region of alpha A-, and alpha B-crystallins in human crystalline lens during aging. *Biochem Biophys Res Commun* 231: 373–378
- Khazaeli AA, Tatar M, Pletcher SD et al. (1997) Heat-induced longevity extension in *Drosophila*. 1. Heat treatment, mortality, and thermotolerance. *J Gerontol A* 52: B48–B52
- Kimura Y, Sakai T, Takeuchi M et al. (1996) An unique CD4⁺CD8⁺ intestinal lymphocyte specific for DnaK (*Escherichia coli* Hsp70) may be selected by intestinal microflora of rats. *Immunobiology* 196: 550–556
- Krawczyk Z and Szymik N (1989) Effect of age and busulphan treatment of the hsp70 gene-related transcript level in rat testes. *Int J Androl* 12: 72–79
- Kregel KC and Moseley PL (1996) Differential effects of exercise and heat stress on liver hsp70 accumulation with aging. *J Appl Physiol* 80: 547–551
- Lin P, Smith DL and Smith JB (1997) *In vivo* modification of the C-terminal lysine of human lens alphaB-crystallin. *Exp Eye Res* 65: 673–680
- Lithgow GJ and Kirkwood TBL (1996) Mechanisms and evolution of aging. *Science* 273: 80
- Lithgow GJ, White T, Melov S et al. (1995) Thermotolerance and extended life-span conferred by single-gene mutations and induced by thermal stress. *Proc Natl Acad Sci USA* 92: 7540–7544

- Liu AY-C, Lin Z, Choi H-S et al. (1989) Attenuated induction of heat shock gene expression in aging diploid fibroblasts. *J Biol Chem* 264: 12037–12045
- Liu AY-C, Lee Y-K, Manalo D et al. (1996) Attenuated heat shock transcriptional response in aging: molecular mechanism and implication in the biology of aging. In: Feige U, Morimoto RI, Yahara I et al. (eds) *Stress Inducible Cellular Responses*. EXS Vol 77, pp 393–408. Birkhauser Verlag, Basel
- Locke M and Tanguay RM (1996) Diminished heat shock response in the aged myocardium. *Cell Stress and Chaperones* 1: 251–260
- Maiello M, Boeri D, Sampietro L et al. (1998) Basal synthesis of heat shock protein 70 increases with age in rat kidneys. *Gerontology* 44: 15–20
- Marchalonis JJ, Schluter SF, Wilson L et al. (1993) Natural human antibodies to synthetic peptide autoantigens: correlation with age and autoimmune disease. *Gerontology* 39: 65–79
- Miyaishi O, Ito Y, Kozaki K et al. (1995) Age-related attenuation of Hsp47 heat response in fibroblasts. *Mech Ageing Dev* 77: 213–226
- Monnier VM and Cerami A (1981) Nonenzymatic browning *in vivo*: possible process for aging of long-lived proteins. *Science* 211: 491–493
- Moore SA, Lopez A, Richardson A et al. (1998) Effect of age and dietary restriction on expression of heat shock protein 70 in rat alveolar macrophages. *Mech Ageing Dev* 104: 59–73
- Morimoto RI (1998) Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones, and negative regulators. *Genes Dev* 12: 3788–3796
- Nakanishi Y and Yasumoto K (1997) Induction after administering paraquat of heme oxygenase-1 and heat shock protein in the liver of senescence-accelerated mice. *Biosci Biotechnol Biochem* 61: 1302–1306
- Niedzwiecki A, Kongpachith AM and Fleming JE (1991) Aging affects expression of 70-kDa heat shock proteins in *Drosophila*. *J Biol Chem* 266: 9332–9338
- Niedzwiecki A, Reveillaud I and Fleming JE (1992) Changing in superoxide dismutase and catalase in aging heat-shocked *Drosophila*. *Free Radic Res Commun* 17: 355–367
- Nitta Y, Abe K, Aoki M et al. (1994) Diminished heat shock protein 70 mRNA induction in aged rat hearts after ischemia. *Am J Physiol* 267: H1795–H1803
- Pahlavani MA, Harris MD, Moore SA et al. (1995) The expression of heat shock protein 70 decreases with age in lymphocytes from rats and rhesus monkeys. *Exp Cell Res* 218: 310–318
- Pahlavani MA, Harris MD, Moore SA et al. (1996) Expression of heat shock protein 70 in rat spleen lymphocytes is affected by age but not by food restriction. *J Nutr* 126: 2069–2075
- Perez N, Sugar J, Charya S et al. (1991) Increased synthesis and accumulation of heat shock 70 proteins in Alzheimer's disease. *Brain Res Mol Brain Res* 11: 249–254
- Piotrowicz RS, Weber LA, Hickey E et al. (1995) Accelerated growth and senescence of arterial cells expressing the small molecular weight heat-shock protein Hsp27. *FASEB J* 9: 1079–1084
- Portig I, Pankuweit S and Maisch B (1997) Antibodies against stress proteins in sera of patients with dilated cardiomyopathy. *J Mol Cell Cardiol* 29: 2245–2251
- Pratt WB (1997) The role of the hsp90-based chaperone system in signal transduction by nuclear receptors and receptors signaling *via* MAP kinase. *Annu Rev Pharmacol Toxicol* 37: 297–326
- Pratt WB and Toft DO (1997) Steroid receptor interactions with heat shock protein and immunophilin complexes. *Endocrine Rev* 18: 306–360
- Rao DV, Watson K and Jones GL (1999) Age-related attenuation in the expression of the major heat shock proteins in human peripheral lymphocytes. *Mech Ageing Dev* 107: 105–118
- Razzaque MS, Shimokawa I, Nazneen A et al. (1998) Age-related nephropathy in the Fischer 344 rat is associated with overexpression of collagens and collagen-binding heat shock protein 47. *Cell Tissue Res* 293: 471–478
- Reiss U and Rothstein M (1974) Heat-labile isozymes of isocitrate lyase from aging *Turbatrix aceti*. *Biochem Biophys Res Commun* 61: 1012–1016
- Renkawek K, Bosman GJ and Gaestel M (1993) Increased expression of heat-shock protein 27 kDa in Alzheimer disease: a preliminary study. *Neuroreport* 5: 14–16
- Rutherford SL and Lindquist S (1998) Hsp90 as a capacitor for morphological evolution. *Nature* 396: 336–342
- Schauer JA and Gafni A (1995) Photodegradation of tryptophan residues and attenuation of molecular chaperone activity in alpha-crystallin are correlated. *Biochem Biophys Res Commun* 212: 900–905
- Schirmer EC, Glover JR, Singer MA et al. (1996) HSP100/Clp proteins: a common mechanism explains diverse functions. *Trends Biochem Sci* 21: 289–296
- Schliwa M, van Blerkom J and Porter KR (1981) Stabilization of the cytoplasmic ground substance in detergent-opened cells and a structural and biochemical analysis of its composition. *Proc Natl Acad Sci USA* 78: 4329–4333
- Serrano M, Lin AW, McCurrah ME et al. (1997) Oncogenic *ras* provokes premature cell senescence associated with accumulation of p53 and p16^{INK4a}. *Cell* 88: 593–602
- Shama S, Lai CY, Antoniazzi JM et al. (1998) Heat stress-induced life span extension in yeast. *Exp Cell Res* 245: 379–388
- Sharma KK and Ortwerth BJ (1995) Effect of cross-linking on the chaperone-like function of alpha crystallin. *Exp Eye Res* 61: 413–421
- Shepherd JC, Walldorf U, Hug P et al. (1989) Fruit flies with additional expression of the elongation factor EF-1 alpha live longer. *Proc Natl Acad Sci USA* 86: 7520–7521
- Shikama N, Ackermann R and Brack C (1994) Protein synthesis elongation factor EF-1 alpha expression and longevity in *Drosophila melanogaster*. *Proc Natl Acad Sci USA* 91: 4199–4203
- Shinohara H, Inaguma Y, Goto S et al. (1993) Alpha B crystallin and Hsp28 are enhanced in the cerebral cortex of patients with Alzheimer's disease. *J Neurol Sci* 119: 203–208
- Smith JB, Jiang X and Abraham EC (1997) Identification of hydrogen peroxide oxidation sites of alpha A- and alpha B-crystallins. *Free Radic Res* 26: 103–111
- Smith JR and Pereira-Smith OM (1996) Replicative senescence: implications for *in vivo* aging and tumor suppression. *Science* 273: 63–67
- Sohal RS and Weindruch R (1996) Oxidative stress, caloric restriction, and aging. *Science* 273: 59–63
- Stadtman ER (1992) Protein oxidation and aging. *Science* 257: 1220–1224
- Sun J, Kale SP, Childress AM et al. (1994) Divergent roles of *RAS1* and *RAS2* in yeast longevity. *J Biol Chem* 269: 18638–18645
- Sun H, Gao J, Ferrington DA et al. (1999) Repair of oxidized calmodulin by methionine sulfoxide reductase restores ability to activate the plasma membrane Ca-ATPase. *Biochemistry* 38: 105–112
- Tatar M, Khazaeli AA and Curtsinger JW (1997) Chaperoning extended life. *Nature* 390: 30
- Voorter CEM, de Haard-Hoekman WA, van den Oetelaar PJM et al. (1988) Spontaneous peptide bond cleavage in aging α -crystallin

- through a succinimide intermediate. *J Biol Chem* 263: 19020–19023
- Wadhwa R, Kaul SC, Ikawa Y et al. (1993a) Identification of a novel member of mouse hsp70 family. Its association with cellular mortal phenotype. *J Biol Chem* 268: 6615–6621
- Wadhwa R, Kaul SC, Sugimoto Y et al. (1993b) Induction of cellular senescence by transfection of cytosolic mortalin cDNA in NIH 3T3 cells. *J Biol Chem* 268: 22239–22242
- Wakutani Y, Urakami K, Shimomura T et al. (1995) Heat shock protein 70 mRNA levels in mononuclear blood cells from patients with dementia of the Alzheimer type. *Dementia* 6: 301–305
- Wheeler JC, Bieschke ET and Tower J (1995) Muscle-specific expression of *Drosophila* hsp70 in response to aging and oxidative stress. *Proc Natl Acad Sci USA* 92: 10408–10412
- Wolosewick JJ and Porter KR (1979) Microtrabecular lattice of the cytoplasmic ground substance. Artifact or reality. *J Cell Biol* 82: 114–139
- Wright HT (1991) Nonenzymatic deamination of asparaginyl and glutaminyl residues in proteins. *Crit Rev Biochem Mol Biol* 26: 1–52
- Wu B, Gu MJ, Heydari AR et al. (1993) The effect of age on the synthesis of two heat shock proteins in the hsp70 family. *J Gerontol* 48: B50–B56
- Youngman LD, Park J-YK and Ames BN (1992) Protein oxidation associated with aging is reduced by dietary restriction of protein or calories. *Proc Natl Acad Sci USA* 89: 9112–9116
- Yuh KCM and Gafni A (1987) Reversal of age-related effects in rat muscle phosphoglycerate kinase. *Proc Natl Acad Sci USA* 84: 7458–7462

