

Spondyloarthropathies and bone resorption: A possible role of heat shock protein (Hsp70) (Review)

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Spondyloarthropathies consist of chronic inflammatory disorders genetically linked with each other through HLA-B27 molecules, and are connected with the destruction of periarticular bone and also with systemic bone loss in many cases. Expected molecular mechanisms behind these conditions overlap the functions of Hsp70s, a group of major molecular chaperones and cytokines. Hsp70s may control disease progression via inhibition of unfolded HLA-B27 protein accumulation and alteration of ER stress signaling. Further, Hsp70s may improve disease related malfunction of antigen presentation, and may induce nitric oxide (NO) release from macrophages which probably protective against spondyloarthropathies as well. Considering premised possible influence of Hsp70s on core mechanisms of spondyloarthropathies it may be expected that, increased expression of Hsp70s advantageously retards disease progress, or may lead to remission. On the other hand Hsp70s as danger signals induces the secretion of proinflammatory cytokines playing major role in the progression of spondyloarthropathy induced bone loss. Consequently, the effect of Hsp70s on the progression of spondyloarthropathic bone loss is “Janus-faced” in some respect: increase of Hsp70s’ level is likely advantageous regarding to the core of disorder; but it may facilitate existing bone resorption processes.

Keywords: antigen presentation, bone resorption, chaperokine, ER stress, heat shock proteins, HLA-B27, Hsp70, osteoblast, osteoclast, spondyloarthropathy

The disease group of spondyloarthropathies consists of several chronic inflammatory disorders such as ankylosing spondylitis, psoriatic spondylitis, inflammatory bowel disease-related spondyloarthritis and rheumatoid arthritis (28, 29). All premised disorders are genetically linked with each other through a human leukocyte antigen B27 type

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molecule (HLA-B27) of type-1 major histocompatibility complex (MHC-I) (28, 47, 48), and are connected with the destruction of periarticular bone and also with systemic bone loss in many cases (26, 29). The consequences of bone resorption may include joint destruction, increased bone fracture risk, implant failures and accelerated loss of periodontal (alveolar) bone (6, 29, 50) causing strong difficulties for several medical and dental professionals and their patients. Accordingly, the link between spondyloarthropathy induced inflammation and consequent bone loss has achieved increasing attention over the past decade (28, 29). Growing research on the molecular mechanisms involved in spondyloarthropathies and consequent inflammation induced changes of bone metabolism has revealed a set of expected molecular mechanisms behind premised pathological processes (28, 29). Interestingly, many of expected molecular mechanism overlap the function of 70 kDa heat shock proteins (Hsp70s) which are major molecular chaperones and cytokines/chaperokines (4) of most cells/tissues (14, 15, 17), extracellular/interstitial fluids (12), blood (43, 58), synovial fluids (37, 49) and also secretory body fluids like saliva (19, 20, 21). Although premised overlap of functions and functional targets is obvious, the possible role of Hsp70s in the pathogenesis of spondyloarthropathies related bone loss was not yet pointed out or discussed in detail. The aim of present review is to stop premised gap of related literature, and to highlight the possible role of Hsp70s in premised pathological processes.

Spondyloarthropathies and the HLA-B27 molecules

Although spondyloarthropathies are known as multifactor disorders; genetic factors are also expected being a major determinant of these disorders (11, 28). Importantly, strong association between spondyloarthropathies and expression of HLA-B27 molecules was clearly confirmed during the past decades (28, 52). Two sets of HLA-B27 related theories are currently being proposed such as antigen specific theories and theories independent of antigen specificity (28). *Antigen specific theories* either predict that the HLA-B27 molecule has a unique ability to bind (and present) joint-specific peptide(s) recognized by autoreactive CD8⁺ T cells (cluster differentiation 8⁺ lymphocytes of thymus origin; cytotoxic T cells) responsible for inflammatory disease; or based on cross-reactivity between some bacterial antigens and HLA-B27 leading to inflammatory disease (28, 30, 38). In latter case other cross reactive self-peptides presented by HLA-B27 itself may also be expected (2, 7, 28). *Theories independent of antigen specificity* are based on unusual biochemical properties of the HLA-B27 type molecules characterized by slow folding during its three-dimensional formation, and tendency to dimerize via “aberrant” intermolecular disulfide bonding (16). Premised properties may lead to misfolding and accumulation of HLA-B27 molecules in the endoplasmic reticulum /ER/ (16). Therefore, these theories expect either proinflammatory ER stress response because of unfolded protein accumulation; or *abnormal* reactivity of HLA-B27 dimers with the receptors of CD4⁺ T cells (cluster differentiation 4⁺ lymphocytes of thymus origin; T helper cells) and natural killer (NK) cells (3, 8, 28). Although both

premised major sets of HLA-B27 related theories underlie the role of T lymphocytes (i.e. CD4⁺ and/or CD8⁺ cells) in the pathomechanism; *malfunction of HLA-B27 coupled antigen presentation* together with *normally functioning T cell response* seems to be the crucial point of disease predisposition (9, 27, 28, 39).

The important role of microbial flora in the development of spondyloarthropathies was also demonstrated via germ-free animal experiments (28, 45, 51). It may be expected that, above mentioned malfunction of antigen presentation impairs immune defense and CD8⁺ cytotoxic response against microbes (22, 23, 55) which may lead to consequent over-activation (“rebound”) of the immune system *in time* leading to impairment of tolerance, particularly towards the microbial flora (27) and cross reactive self-peptides. Further, the possible role of several inflammatory mediators such as interferon gamma (INF- γ), interleukin 2 (IL-2) and also nitric oxide (NO) was also expected (10, 28). The possible role of INF- γ and IL-2 in the pathomechanism is far from being clear, but protective role of nitric oxide (NO) against spondyloarthropathies is likely (10, 28).

Bone loss and spondyloarthropathies

Osteoclast formation and consequent disturbance of the tight balance between bone resorption and bone formation seems to be an essential step in chronic inflammatory bone resorption induced by spondyloarthropathies (29, 44). This unfavorable shift of balance towards bone resorption is the basis for rapid bone loss primarily (29). There could be several mechanisms expected behind osteoclast activation, however the interrelationship between the immune system and the differentiation of osteoclasts seems to be a rather important and determinant one (29). Premised interrelationship may be routed in the fact that, immune cells and osteoclast precursor cells are derived from the same hematopoietic precursor cells (whereas osteoblast precursors are cells of mesenchymal origin) (29). Consequently, pro-inflammatory cytokine signaling network responsible for the activation of immune cells exert an activating effect on osteoclast precursor cells (having the same origin like immune cells) rather than on osteoblast precursors; which lead to the shift towards bone resorption (29). Further, certain pro-inflammatory cytokines, such as interleukins (IL) IL-1 β (56), IL-6 (57), IL-17 (34) and especially tumor necrosis factor alpha (TNF- α) (36) induce the expression of RANKL (receptor activator of nuclear factor- $\kappa\beta$ ligand; a member of tumor necrosis factor superfamily) which is essential for final differentiation steps of osteoclasts as well as for their bone resorbing capacity (33, 35). TNF- α additionally induces the mobilization of osteoclast precursor cells (OCPs) from the bone marrow and their homing and migration to the sites of inflammation (29). Moreover, TNF- α also exerts inhibitory effects on osteoblasts via degradation of runt-related transcription factor 2 (Runx2) (32) and inhibition of Wnt (a highly conserved signaling molecule, with a name derived from the first two recognized members of the family) induced osteoblast differentiation (18, 31); which further increases the shift toward bone resorption (29).

Spondyloarthropathies, bone loss and Hsp70s

Until now there have been described 25 allelic subtypes coding for 23 distinct HLA-B27 molecules (HLA-B2701 to HLA-B2723), and most subtypes have been disease-associated (28). However, those which seem not to be associated with spondyloarthropathies /such as HLA-B2706 and HLA-B2709 (46)/differ from the others because of amino-acid substitution in position 116 which is involved in the association of HLA-B27 type molecules with chaperones (28) likely including also Hsp70s (41, 54). Considering the antigen specificity independent theories of spondyloarthropathies (see above), and based on premised important finding and known chaperoning and signaling functions of Hsp70 in the ER (40) it is likely that, Hsp70s may play crucial role in controlling disease progression (via inhibition of unfolded HLA-B27 protein accumulation and alteration of ER signaling in response to ER stress). Further, antigen-complexed extracellular Hsp70s enhance the function of antigen presenting cells (i.e., macrophages, dendritic cells) and (cross)presentation of antigens (coupled with either MHC-I or MHC-II molecules) to cytotoxic T cells (CD8+ cells) or T helper cells (CD4+ cells), respectively (12, 53). Although direct links between bone loss and HLA-B27 related ER alterations and/or malfunction of antigen presentation were not yet discussed; it is very likely that premised expected effects of Hsp70s on the core of disease progression decreases the bone-related consequences of spondyloarthropathies as well. Further, uncomplexed extracellular Hsp70s also induce inducible nitric oxide (NO) synthase and NO release from macrophages (42); which is likely to be protective against spondyloarthropathies as well (10, 28). Considering premised possible influence of Hsp70s on core mechanisms of spondyloarthropathies it may be expected that, increase of intra and/or extracellular level of Hsp70s advantageously influences (retards) disease progress, or may lead to remission. It may not be excluded either that, certain effects – like ionic strength, warm or massage etc. (1, 24, 25) – used for physiotherapies may lead to improvement of spondyloarthropathic symptoms due to the increase of intra- (1, 25) and/or extracellular (19, 20, 24) Hsp70s. On the other hand, uncomplexed extracellular Hsp70s as danger signal induces the secretion of proinflammatory cytokines playing major role in the progression of spondyloarthropathy induced bone loss, such as IL-1 β , IL6 and TNF- α (5, 13) as detailed above. Consequently, the effect of Hsp70s on the progression of spondyloarthropathic bone loss is “Janus-faced” in some respect: Increase of Hsp70 level is seems to be advantageous regarding to the core of disorder; but it may facilitate existing bone resorption processes.

Conclusion

Taking together data and considerations above it may be concluded that, Hsp70s likely play an important role in the pathomechanism of spondyloarthropathies and consequent bone loss. Therefore, it would be worth investigating the possible role of intra- and extracellular Hsp70s in the pathomechanism of (as well as possible therapeutic targets for) spondyloarthropathies and related bone loss in detail.

REFERENCES

1. Ait-Aïssa S, Porcher JM, Arrigo AP, Lambré C: Activation of the Hsp70 promoter by environmental inorganic and organic chemicals: relationships with cytotoxicity and lipophilicity. *Toxicology* 145, 147–157 (2000)
2. Allen RL, Bowness P, McMichael AJ: The role of HLA-B27 in spondyloarthritis. *Immunogenetics* 50, 220–227 (1999)
3. Allen RL, Raine T, Haude A, Trowsdaale J, Wilson MJ: Leukocyte receptor complex-encoded immunomodulatory receptors show differing specificity for alternative HLA-B27 structures. *J. Immunol.* 167, 5543–5547 (2001)
4. Asea A: Stress proteins and initiation of immune response: Chaperokine activity of Hsp72. *Exerc. Immunol. Rev.* 11, 34–35 (2005)
5. Asea A, Kraeft S, Kurt-Jones EA, Stevenson MA, Chen LB, Finberg RW, Koo GC, Calderwood SK: Hsp70 stimulates cytokine production through a CD14 dependent pathway, demonstrating its dual role as a chaperone and cytokine. *Nat. Med.* 6, 435–442 (2000)
6. Barker D, Nohl FS, Postlethwaite KR, Smith DG: Case report of multiple implant failure in a patient with ankylosing spondylitis. *Eur. J. Prosthodont. Restor. Dent.* 16, 20–23 (2008)
7. Boyle LH, Gaston JSH: Breaking the rules: the unconventional recognition of HLA-B27 by CD4+T lymphocytes as an insight into the pathogenesis of the spondyloarthropathies. *Rheumatol.* 41, 1280–1285 (2003)
8. Boyle LH, Goodall JC, Opat SS, Gaston JSH: The recognition of HLA-B27 by human CD4+ T lymphocytes. *J. Immunol.* 167, 2619–2624 (2001)
9. Breban M, Hammer RE, Richardson JA, Taurog JD: Transfer of the inflammatory disease of HLA-B27 transgenic rats by bone marrow engraftment. *J. Exp. Med.* 178, 1607–1616 (1993)
10. Breban M, May E: Treatment of SpA-like disease in HLA-B27 transgenic animals. *Clin. Exp. Rheum.* 20, S50–S51 (2002)
11. Breban M, Said-Nahal R, Hugot JP, Micelli-Richard C: Familial and genetic aspects of spondyloarthropathy. *Rheum. Dis. Clin. North. Am.* 29, 575–594 (2003)
12. Calderwood SK: Extracellular heat shock proteins in cell signaling. *FEBS Letters* 581, 3689–3694 (2007)
13. Campisi J, Fleshner M: The role of extracellular Hsp72 in acute stress-induced potentiation of innate immunity in physically active rats. *J. Appl. Physiol.* 94, 43–52 (2003)
14. Csermely, P: Proteins, RNA-s and chaperones in enzyme evolution: a folding perspective. *Trends Biochem. Sci.* 22, 147–149 (1997)
15. Csermely, P: Chaperone-percolator model: a possible molecular mechanism of Anfinsen-cage-type chaperones. *BioEssay* 21, 959–965 (1999)
16. Dangoria NS, DeLay ML, Kingsbury DJ, Mear JP, Uchanska-Ziegler B, Ziegler A, Colbert RA: HLA-B27 misfolding is associated with aberrant intermolecular disulfide bond formation (dimerization) in the endoplasmic reticulum. *J. Biol. Chem.* 277, 23459–23468 (2002)
17. Daugaard M, Rohde M, Jäättelä M: The heat shock protein 70 family: Highly homologous proteins with overlapping and distinct functions. *FEBS Letters* 581, 3702–3710 (2007)
18. Diarra D, Stolina M, Polzer K, Zwerina J, Ominsky MS, Dwyer D, Korb A, Smolen J, Hoffmann M, Scheinecker C, van der Heide D, Landewe R, Lacey D, Richards WG, Schett G: Dickkopf-1 is a master regulator of joint remodeling. *Nat. Med.* 13, 156–163 (2007)
19. Fábíán TK, Gáspár J, Fejérdy L, Kaán B, Bálint M, Csermely P, Fejérdy P: Hsp70 is present in human saliva. *Med. Sci. Monit.* 9, BR62–BR65 (2003)
20. Fábíán TK, Tóth Zs, Fejérdy L, Kaán B, Csermely P, Fejérdy P: Photo-acoustic stimulation increases the amount of 70 kDa heat shock protein (Hsp70) in human whole saliva. A pilot study. *Int. J. Psychophysiol.* 52, 211–216 (2004)
21. Fábíán TK, Fejérdy P, Nguyen MT, Söti Cs, Csermely P: Potential immunological functions of salivary Hsp70 in mucosal and periodontal defense mechanisms. *Arch. Immunol. Ther. Exp.* 55, 91–98 (2007)
22. Falgarone G, Blanchard HS, Riot B, Simonet M, Breban M: The cytotoxic T cell-mediated response against *Yersenia pseudotuberculosis* in HLA-B27 transgenic rat. *Infect. Immun.* 67, 3773–3779 (1999)

23. Falgarone G, Blanchard HS, Virecoulon F, Simonet M, Breban M: Coordinate involvement of invasin and Yop proteins in a *Yersinia pseudotuberculosis*-specific class I-restricted cytotoxic T cell-mediated response. *J. Immunol.* 162, 2875–2883 (1999)
24. Fejérdy L, Tóth Zs, Kaán B, Fábíán TK, Csermely P, Fejérdy P: Lokális hő- és mechanikai stress (masszázs) stimuláció hatása az emberi kevert nyál molekuláris chaperon (Hsp70) koncentrációjára. Előzetes vizsgálatok. In Hungarian (The effect of heat stimulation and mechanical stress (massage) of salivary glands on the secretoric parameters of salivary Hsp70. A pilot study) *Fogorv. Szle.* 97, 204–210 (2004)
25. Grasso S, Sciciffo C, Cardille V, Gulino R, Renis M: Adaptive response to the stress induced by hyperthermia or hydrogen peroxide in human fibroblasts. *Exp. Biol. Med.* 228, 491–198 (2003)
26. Gough AK, Lilley J, Eyre S, Holder RL, Emery P: Generalized bone loss in patients with early rheumatoid arthritis. *Lancet* 344, 23–27 (1994)
27. Hacquard-Bouder C, Falgarone G, Bosquet A, Smaoui F, Monnet D, Ittah M, Breban M: Defective costimulatory function is a striking feature of antigen-presenting cells in an HLA-B27-transgenic rat model of spondyloarthropathy. *Arthritis. Rheum.* 50, 1624–1635 (2004)
28. Hacquard-Bouder C, Ittah M, Breban M: Animal models of HLA-B27-associated diseases: new outcomes. *Joint. Bone. Spine.* 73, 132–138 (2006)
29. Herman S, Krönke G, Schett G: Molecular mechanisms of inflammatory bone damage: emerging targets for therapy. *Trends. Mol. Med.* 14, 245–253 (2008)
30. Jardetzky TS, Lane WS, Robinson RA, Madden DR, Wiley DC: Identification of self peptides bound to purified HLA-B27. *Nature* 353, 326–329 (1991)
31. Johnson KL, Kamel MA: The Wnt signaling pathway and bone metabolism. *Curr. Opin. Rheumatol.* 19, 376–382 (2007)
32. Kaneki H, Guo R, Chen D, Yao Z, Schwarz EM, Zhang YE, Boyce BF, Xing L: Tumor necrosis factor promotes Runx2 degradation through up-regulation of Smurf1 and Smurf2 in osteoblasts. *J. Biol. Chem.* 281, 4326–4333 (2006)
33. Kong YY, Yoshida H, Sarosi I, Tan HL, Timms E, Capparelli C, Morony S, Olivera-dos-Santos AJ, Van G, Itie A, Khoo W, Wakeham A, Dunstan CR, Lacey DL, Mak TW, Boyle WJ, Penninger JM: OPGL is a key regulator of osteoclastogenesis, lymphocyte development and lymph-node organogenesis. *Nature* 397, 315–323 (1999)
34. Kotake S, Udagawa N, Takahashi N, Matsuzaki K, Itoh K, Ishiyama S, Saito S, Inoue K, Kamatani N, Gillespie MT, Martin TJ, Suda T: IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J. Clin. Invest.* 103, 1345–1352 (1999)
35. Lacey DL, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, Elliott R, Colombero A, Elliott G, Scully S, Hsu H, Sullivan J, Hawkins N, Davy E, Capparelli C, Eli A, Qian YX, Kaufman S, Sarosi I, Shalhoub V, Senaldi G, Guo J, Delaney J, Boyle WJ: Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell* 93, 165–176 (1998)
36. Lam J, Takeshita S, Barker JE, Kanagawa O, Ross FP, Teitelbaum SL: TNF-alpha induces osteoclastogenesis by direct stimulation of macrophages exposed to permissive levels of RANK ligand. *J. Clin. Invest.* 106, 1481–1488 (2000)
37. Luo X, Zuo X, Zhang B, Song L, Wei X, Zhou Y, Xiao X: Release of heat shock protein 70 and the effects of extracellular heat shock protein 70 on the production of IL-10 in fibroblast-like synoviocytes. *Cell Stress Chaperones* 13, 365–373 (2008)
38. Madden DR, Gorga JC, Strominger JL, Wiley DC: The structure of HLA-B27 reveals nonamer self-peptides bound in an extended conformation. *Nature* 353, 321–325 (1991)
39. May E, Dorris ML, Satumtira N, Iqbal I, Rehman MI, Lightfoot E, Taurog JD: CD8 alpha beta T cells are not essential to the pathogenesis of arthritis or colitis in HLA-B27 transgenic rats. *J. Immunol.* 170, 1099–1105 (2003)
40. Ni M, Lee AS: ER chaperones in mammalian development and human diseases. *FEBS Letters* 581, 3641–3651 (2007)
41. Nössner E, Parham P: Species – specific differences in chaperone interaction of human mouse major histocompatibility complex class I molecules. *J. Exp. Med.* 181, 327–337 (1995)

42. Panjwani NN, Popova L, Srivastava PK: Heat shock proteins gp96 and hsp70 activate the release of nitric oxide by APCs. *J. Immunol.* 168, 2997–3003 (2002)
43. Rea, IM; McNerlan, S; Pockley, AG. Serum heat shock protein and anti-heat shock protein antibody levels in aging. *Exp. Gerontol.* 2001 36, 341–352.
44. Redlich K, Hayer S, Ricci R, David JP, Tohidast-Akrad M, Kollias G, Steiner G, Smolen JS, Wagner EF, Schett G: Osteoclasts are essential for TNF-alpha-mediated joint destruction. *J. Clin. Invest.* 110, 1419–1427 (2002)
45. Reháková Z, Capková J, Stepanková R, Sinkora J, Louzecká A, Ivanyi P, Weinreich S: Germ-free mice do not develop ankylosing enthesopathy, a spontaneous joint disease. *Hum. Immunol.* 61, 555–558 (2000)
46. Reveille JD, Ball EJ, Khan MA: HLA-B27 and genetic predisposing factors in spondyloarthropathies. *Curr. Opin. Rheumatol.* 13, 265–272 (2001)
47. Said-Nahal R, Miceli-Richard C, Berthelot J-M, Duché A, Dermis-Labous E, Le Blévec G, Saraux A, Perdriger A, Guis S, Claudepierre P, Sibilia J, Amor B, Dougados M, Breban M: The familiar form of spondylarthropathy: a clinical study of 115 multiplex families. *Arthritis Rheum.* 43, 1356–1365 (2000)
48. Said-Nahal R, Miceli-Richard C, D'Agostino MA, Dermis-Labous E, Berthelot J-M, Duché A, Le Blévec G, Saraux A, Perdriger A, Guis S, Amor B, Dougados M, Breban M, Groupe Français d'Etude Génétique des Spondylarthropathies: Phenotypic diversity is not determined by independent genetic factors in familial spondylarthropathy. *Arthritis Rheum.* 45, 478–484 (2001)
49. Suzuki T, Segami N, Nishimura M, Hattori H, Nojima T: Analysis of 70Kd heat shock protein expression in patients with internal derangement of the temporomandibular joint. *Int. J. Oral. Maxillofac. Surg.* 29, 301–304 (2000)
50. Tatakis DN, Guglielmoni P: HLA-B27 transgenic rats are susceptible to accelerated alveolar bone loss. *J. Periodontol.* 71, 1395–1400 (2000)
51. Taurog JD, Richardson JA, Croft JT, Simmons WA, Zhou M, Fernandez-Sueiro JL, Balish E, Hammer RE: The germfree state prevents development of gut and joint inflammatory disease in HLA-B27 transgenic rats. *J. Exp. Med.* 180, 2359–2364 (1994)
52. Taurog JD, Maika S, Satumtira N, Dorris ML, McLean IL, Yanagisawa H, Sayad A, Stagg AJ, Fox GM, Lé O'Brien A, Rehman M, Zhou M, Weiner AL, Splawski JB, Richardson JA, Hammer RE: Inflammatory disease in HLA-B27 transgenic rats. *Immunol. Rev.* 169, 209–223 (1999)
53. Tobian AA, Canaday DH, Harding CV: Bacterial heat shock proteins enhance class II MHC antigen processing and presentation of chaperoned peptides to CD4+ T cells. *J. Immunol.* 173, 5130–5137 (2004)
54. Tran TM, Satumtira N, Dorris ML, May E, Wang A, Furuta E, Taurog JD: HLA-B27 in transgenic rats forms disulfide – linked heavy chain oligomers and multimers that bind to the chaperone BiP. *J. Immunol.* 172, 5110–5119 (2004)
55. Warner TF, Madsen J, Starling J, Wagner RD, Taurog JD, Balish E: Human HLA-B27 gene enhances susceptibility of rats to oral infection by *Listeria monocytogenes*. *Am. J. Pathol.* 149, 1737–1743 (1996)
56. Wei S, Kitaura H, Zhou P, Ross FP, Teitelbaum SL: IL-1 mediates TNF-induced osteoclastogenesis. *J. Clin. Invest.* 2005 115, 282–290 (2005)
57. Wong PK, Quinn JM, Sims NA, van Nieuwenhuijze A, Campbell IK, Wicks IP: Interleukin-6 modulates production of T-lymphocyte-derived cytokines in antigen-induced arthritis and drives inflammation-induced osteoclastogenesis. *Arthritis Rheum.* 48, 158–168 (2006)
58. Wright BH, Corton JM, El-Nahas AM, Wood RF, Pockley AG: Elevated levels of circulating heat shock protein 70 (Hsp70) in peripheral and renal vascular disease. *Heart Vessels* 15, 18–22 (2000)